

Supplementary Information for
**VinaMPI: Facilitating Multiple Receptor High-Throughput Virtual Docking on High
Performance Computers**

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VinaMPI Tutorial

Compiling VinaMPI (these directions were used on the Kraken supercomputer)

1. Download the revised Autodock Vina code and VinaMPI (install directory = \$INSTALL_DIR)

```
~> cd $INSTALL_DIR
~> wget cmb.ornl.gov/~sek/autodock_vina_1_1_2.tgz
~> wget cmb.ornl.gov/~sek/VinaMPIv2.tgz
~> tar xzvf autodock_vina_1_1_2.tgz
~> tar xzvf VinaMPIv2.tgz
```

2. Compile the Autodock Vina code

```
~> cd autodock_vina_1_1_2_rev/autodock_vina_1_1_2/build/linux/release/
~> module swap PrgEnv-pgi PrgEnv-gnu
~> module load boost
~> make clean
~> make depend (this step may give warnings but still be ok)
~> make
```

3. Direct VinaMPI to the Autodock Vina code and compile VinaMPI

```
~> cd $INSTALL_DIR/VinaMPIv2_new2/
~> vi vinampi.work.c
```

```
go to line 279 and change the path to Autodock Vina to
$INSTALL_DIR/autodock_vina_1_1_2_rev/autodock_vina_1_1_2/build/linux/release/vi
na
save and quit vi
```

```
~> make
```

File Preparation

1. Changes have been made to a few ADT scripts in order to prepare the input file needed by VinaMPI and additional scripts written to facilitate the process. Download files and copy them to the ADT directories on your computer.

```
~> wget cmb.ornl.gov/~sek/predocking.tgz
~> tar xzvf predocking.tgz
~> mv MoleculePreparation2.py
/Library/MGLTools/1.5.4/MGLToolsPckgs/AutodockTools/
~> mv prepare_ligand5.py
/Library/MGLTools/1.5.4/MGLToolsPckgs/AutodockTools/Utilities24
```

Make sure `predockingflexv4.py` correctly points to `prepare_ligands5.py` and the `parentDirectory` points to the directory containing your input `.mol2` file and lines 30 and 31 have the correct string denoting a new molecule in the `.mol2` file.

2. Starting with a `.mol2` file containing all of the ligands for your screen call the `predockingflexv4.py` scripts. As a parameter it takes the name of the `.mol2` file.

```
~> python predockingflexv4.py input.mol2
```

This will create all the ligand input `.pdbqt` files for the screen and a file `ligands.txt` which contains all the ligand names, number of rotatable bonds, and number of atoms.

3. Call the `preprocess_ligandsv3.py` to sort the `ligands.txt` file and create the input file needed for VinaMPI (`ligand_sort.txt`).

```
~> python preprocess_ligandsv3.py
```

4. Make sure that `prep_receptors.py` points to `prepare_receptor4.py` (ADT scripts) and `parentDirectory` points to directory in which you want the receptor `.pdbqt` files to be created. Then call the script from within the directory containing the `.pdb` files for all of the receptors.

```
~> python prep_receptors.py
```

5. If doing ensemble docking where you are specifying the center of the docking box based on one atom in the protein and your different receptor files are different conformations of the same protein, you can use the `make_rec_param.py` script and give it the atom and size of box side in Å (if cubic) as parameters. If every receptor is a unique protein, the receptor input file must be created manually by the guidelines in the manuscript.

```
~> python make_rec_param.py VAL_281_CG1 30
```

6. Once all of the files are in place, the screen can be started with the following command

```
aprun -n 84672 VinaMPI receptors.txt ligands_sort.txt 4 98164 receptors ligands
```

where `-n` specifies the number of processors to use (flags would be changed here if using threading as well), `VinaMPI` is the path to the program, `receptors.txt` is the path to the receptor parameter file, `ligands_sort.txt` is the path to the ligand parameter file, `4` is the number of receptors in the screen, `98164` is the number of ligands in the screen, `receptors` is the directory containing all of the receptor `.pdbqt` input files, and `ligands` is the directory containing all of the ligand `.pdbqt` input files. The output files will be written in a directory named `out` created in the directory in which the program is called.